



## User's Manual

# Monkey Tetanus Toxoid IgG1 ELISA Kit



DEIASL077



96T





This product is for research use only and is not intended for diagnostic use.

For illustrative purposes only. To perform the assay the instructions for use provided with the kit have to be used.

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## PRODUCT INFORMATION

### Intended Use

This test kit allows rapid and quantitative measurement of antitetanus toxoid IgG1 levels in serum or plasma. IgG1 is the major IgG subclass in monkeys.

### General Description

Drug candidates are routinely screened for evidence of immune system regulation during the discovery process. It is important that the immune response is not enhanced or decreased since such effects may lead to hypersensitivity or increased susceptibility to infection. Determination of a drug candidate's effects on antitetanus toxoid antibody levels allows easy assessment of immune system regulation. Animals are immunized with tetanus toxoid while undergoing drug treatment and serum is collected at appropriate times post immunization. Serum collected 5-7 days after immunization is used for measurement of anti-tetanus toxoid IgM levels, and serum collected 14+ days post immunization is used to measure anti-tetanus toxoid IgG levels. Comparison of anti-tetanus toxoid IgG or IgM levels in drug treated versus control groups reveals effects on the immune response.

### Principles of Testing

The monkey anti-tetanus toxoid IgG test kit is based on a solid phase enzyme-linked immunosorbent assay (ELISA). The assay uses tetanus toxoid for solid phase (microtiter wells) immobilization and horseradish peroxidase (HRP) conjugated mouse monoclonal anti-monkey IgG1 antibody for detection. Serum or plasma samples are diluted and incubated in the microtiter wells for 60 minutes. The microtiter wells are subsequently washed, and HRP conjugate is added and incubated for 45 minutes. Anti-tetanus toxoid IgG1 molecules are thus sandwiched between immobilized tetanus toxoid and the detection antibody conjugate. The wells are then washed to remove unbound HRP-labeled antibodies, and TMB Reagent is added and incubated for 20 minutes at room temperature. This results in the development of a blue color. Color development is stopped by the addition of Stop Solution, changing the color to yellow, and optical density is measured spectrophotometrically at 450 nm. The concentration of antitetanus toxoid IgG1 is proportional to the optical density of the test sample.

### Reagents And Materials Provided

1. Tetanus Toxoid Coated 96-well Plate (12 strips of 8 wells)
2. Enzyme Conjugate Reagent, 11 mL
3. Reference Standard (lyophilized) 2 vials. Store < -20°C
4. 20x Wash Solution, 50 mL
5. Diluent, 30 mL
6. TMB Reagent (One-Step), 11 mL
7. Stop Solution (1N HCl), 11 mL

## Materials Required But Not Supplied

1. Precision pipettes and tips
2. Distilled or deionized water
3. Polypropylene or glass tubes
4. Vortex mixer
5. Absorbent paper or paper towels
6. Micro-plate incubator/shaker with mixing speed of ~150 rpm
7. Plate washer
8. Plate reader with an optical density range of 0-4 at 450nm
9. Graph paper (PC graphing software is optional)

## Storage

1. The reference standard should be stored at -20°C for optimal stability.
2. All remaining kit components should be stored at 2-8°C.
3. The microtiter plate should be kept in a sealed bag with desiccant to minimize exposure to damp air. Test kit will remain stable for six months from the date of purchase provided that the components are stored as described above.

## Specimen Collection And Preparation

General Note: The level of anti-tetanus toxoid IgG1 will depend on dose, route of immunization and time of sample collection. We found that anti-tetanus toxoid IgG1 is present in monkey serum at concentrations of 50 µg/mL or greater. We suggest that samples initially be diluted 200 fold using the following procedure. Optimal dilutions must be determined empirically. Dilutions of 50 fold or lower should not be used.

1. Dispense 298.5 µL of diluent into separate polypropylene or glass tubes.
2. Pipette and mix 1.5 µL of the serum/plasma sample into the tube containing 298.5 µL of diluent. This provides a 200 fold diluted sample.
3. Repeat this procedure for each sample to be tested.

## Reagent Preparation

### WASH SOLUTION PREPARATION:

The wash solution is provided as a 20x stock. Prior to use, dilute the contents of the bottle (50 mL) with 950 mL of distilled or deionized water.

### STANDARD PREPARATION:

1. The monkey anti-tetanus toxoid IgG1 reference standard is provided as a lyophilized stock. Reconstitute with the volume of diluent indicated on the vial label.

2. Prepare a 2.5 µg/mL working standard of monkey anti-tetanus toxoid IgG1 as directed on the reference standard vial label (the reconstituted standard should be frozen at or below -20°C after reconstitution if additional use is intended).
3. Label 5 polypropylene or glass tubes as 1.25, 0.625, 0.313, 0.156 and 0.078 µg/mL and pipette 250 µL of diluent into each tube.
4. Into the tube labeled 1.25 µg/mL, pipette and mix 250 µL of the 2.5 µg/mL anti-tetanus toxoid IgG with 250 µg of diluent. This provides the 1.25 µg/mL standard.
5. Similarly prepare the 0.625, 0.313, 0.156 and 0.078 µg/mL standards by serial dilution.

## Assay Procedure

1. Secure the desired number of coated wells in the holder.
2. Dispense 100 µL of standards (2.5-0.078 µg/mL) and diluted samples into the wells (standards and samples should be tested in duplicate).
3. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 60 minutes.
4. Aspirate the contents of the microtiter wells and wash the wells 5-6 times with 1x wash solution using a plate washer (400µL/well). The entire wash procedure should be performed as quickly as possible.
5. Strike the wells sharply onto absorbent paper or paper towels to remove all residual wash buffer.
6. Add 100 µL of enzyme conjugate reagent into each well.
7. Incubate on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 45 minutes.
8. Wash as detailed in 4 to 5 above.
9. Dispense 100 µL of TMB Reagent into each well.
10. Gently mix on an orbital micro-plate shaker at 100-150 rpm at room temperature (18-25°C) for 20 minutes.
11. Stop the reaction by adding 100 µL of Stop Solution to each well.
12. Gently mix. It is important to make sure that all the blue color changes to yellow.
13. Read the optical density at 450 nm with a microtiter plate reader within 5 minutes.

## Calculation

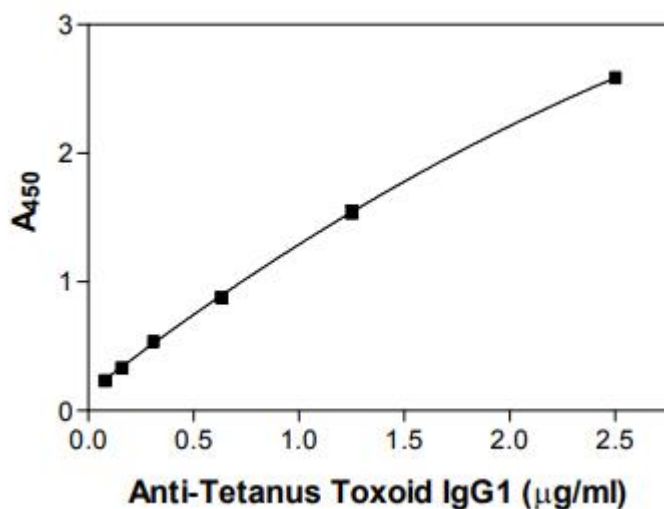
1. Calculate the average absorbance values (A450) for each set of reference standards and samples.
2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in µg/mL on linear graph paper, with absorbance values on the vertical or Y-axis and concentrations on the horizontal or X-axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of anti-tetanus toxoid IgG1 in µg/mL from the standard curve.
4. Multiply the derived concentration by the dilution factor to determine the actual concentration of anti-tetanus toxoid IgG1 in the serum/plasma sample.
5. PC graphing software may be used for the above steps.
6. If the OD450 values of samples fall outside the standard curve when tested at a dilution of 200, samples should be diluted appropriately and re-tested.



## Typical Standard Curve

A typical standard curve with optical density readings at 450 nm on the Y-axis against anti-tetanus toxoid IgG1 concentrations on the X-axis is shown below. This curve is for the purpose of illustration only and should not be used to calculate unknowns. Each user should obtain his or her data and standard curve in each experiment.

Anti-tetanus toxoid IgG1 ( $\mu\text{g/ml}$ )	$A_{450}$
2.500	2.587
1.250	1.543
0.625	0.884
0.313	0.542
0.156	0.336
0.078	0.236



## Precautions

1. Please read and understand the instructions thoroughly before using the kit.
2. All reagents should be allowed to reach room temperature (18-25°C) before use.
3. The assay was designed for use with serum or plasma obtained from monkeys 14 days or more after immunization with tetanus toxoid, at which point the immune response originates predominantly from IgG.
4. The optimal sample dilution should be determined empirically. Do not test samples at dilutions below 50 fold.
5. Optimal results are achieved if, at each step, reagents are pipetted into the wells of the microtiter plate within 5 minutes.

## Limitations

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of and in accordance with the instructions detailed above.
2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.